

HUMAN CXCL14/BRAK ELISA KIT

FOR THE QUANTITATIVE DETERMINATION OF HUMAN CXCL14/BRAK CONCENTRATIONS IN SERUM, PLASMA AND CELL CULTURE SUPERNATES



FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

PURCHASE INFORMATION:

ELISA NAME	HUMAN CXCL14/BRAK ELISA
Catalog No.	SK00439-01
Lot No.	
Formulation	96 T
Standard range	125 - 4000 pg/mL
Sensitivity	62.5 pg/mL
Sample require	100 µL
Dilution Factor	Optimal dilutions should be determined by each laboratory for each application
Sample Type	Serum, Plasma, Cell Culture Supernates
Specificity	Human CXCL14
Intra-assay Precision	6-8%
Inter-assay Precision	8-12%
Storage	2-8°C

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INTRODUCTION

Human CXCL14 immunoassay is a 3.5 - 4.5 hour solid phase ELISA designed to measure Human CXCL14 in cell culture supernates, serum and plasma. It contains recombinant human CXCL14 and antibodies raised against this protein. It has been shown to accurately quantify recombinant human CXCL14. Results obtained with naturally occurring CXCL14 samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural human CXCL14.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for CXCL14 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any CXCL14 present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated antibody specific for CXCL14 is added to the wells. Following a wash to remove any unbound antibody reagent, HRP link Streptavidin is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of CXCL14 bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

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_ The kit should not be used beyond the expiration date on the kit label.

_ Do not mix or substitute reagents with those from other lots or sources.

_ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

_ If samples generate values higher than the highest standard, dilute the samples with Dilution Buffer and repeat the assay.

_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

_ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all

factors have been tested in the immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
CXCL14 Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with a purified antibody against CXCL14.	439-01-01	1 plate
CXCL14 Standard – 4000 pg/vial of recombinant human CXCL14 in a buffered protein base with preservatives; lyophilized.	439-01-02	1 vial
Detection Antibody – 1.05 mL/vial, 10-fold Concentrate of Biotinylated antibody against CXCL14 with preservatives; lyophilized.	439-01-03	1 vial
Positive Control – one vial of recombinant human CXCL14, lyophilized	439-01-04	1 vial
Streptavidin-HRP Conjugate – 60 µL/vial, 200-fold concentrated solution of Streptavidin conjugate to HRP	SAHRP	1 vial
Dilution Buffer – 60 mL of buffered protein based solution with preservatives	DB01	1 bottle
Wash Buffer – 50 mL of 10-fold concentrated buffered surfactant, with preservative.	WB01	1 bottle
TMB Substrate Solution – 11 mL of TMB substrate solution	TMB01	1 bottle
Stop Solution – 11 mL of 0.5M HCl	S-STOP	1 bottle
Plate Sealer	EAPS	1
Plastic Pouch	P01	1

STORAGE

Unopened Kit: Store at 2 - 8° C for up to 12 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should be stored at -20°C or -70°C. Do not use kit past expiration date.

Opened / Reconstituted Reagents: Reconstituted Standard and Detection Antibody Concentrated Solution SHOULD BE STORED at -20°C or -70°C for up to one month. Streptavidin-HRP Conjugate 200-

fold concentrated solution (**protect from light**) and other components may be stored at 2 - 8°C for up to 12 months.

Microplate Wells: Return unused wells to the plastic pouch with the desiccant pack and seal along entire edge of zip-seal. Microplate may be stored for up to 6 months at 2 - 8°C after opening.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

PRECAUTIONS FOR USE

All reagents should be considered as potentially hazardous. The stop solution contains diluted Hydrochloric acid. Appropriate care, therefore, should be taken while handling this solution. We therefore recommend that this product be handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water.

SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤-20° C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤-20°C. Avoid repeated freeze-thaw cycles.

Note: Use Aprotinin (enzyme inhibitor) (Code No.: 00700-01-25) for ALL sample collection to prevent

sample degradation. 0.5 TIU per ml of sample solution.

SAMPLE PREPARATION

Optimal dilutions should be determined by each laboratory for each application.

Use polypropylene test tubes.

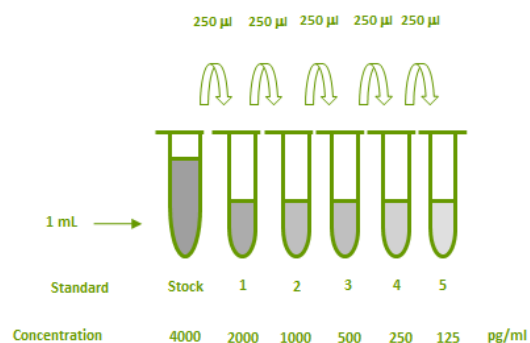
REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of Wash Buffer.

CXCL14 Standard - Refer to vial label for reconstitution volume. Reconstitute the **CXCL14 Standard** with 1.0 mL of Dilution Buffer. This reconstitution produces a stock solution of 4000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 µL of Dilution Buffer into tubes #1 to #5. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The **4000 pg/mL** standard serves as the high standard. The Dilution Buffer serves as the zero standard (0 pg/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
stock	powder	1.0 mL	4000 pg/mL
# 1	250µL of stock	250µL	2000 pg/mL
# 2	250µL of 1	250µL	1000 pg/mL
# 3	250µL of 2	250µL	500 pg/mL
# 4	250µL of 3	250µL	250 pg/mL
# 5	250µL of 4	250µL	125 pg/mL



Detection Antibody - Reconstitute the Detection Antibody Concentrate with 1.05 mL of Dilution

Buffer to produce a 10-fold concentrated stock solution. Pipette 9.45 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 1.05 mL of 10-fold concentrated stock solution to prepare working solution.

Streptavidin-HRP Conjugate - Pipette 11.94 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 60 μ L of 200-fold concentrated stock solution to prepare working solution. *Note: 1x working solution of Streptavidin-HRP Conjugate should be used within a few days. Protect from light.*

Positive Control - Reconstitute the **Positive Control** with 0.5 mL of Dilution Buffer. *Note: Positive Control should be prepared and used immediately. Reconstituted Positive Control CAN NOT BE REUSED.*

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and samples be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess micro-plate strips from the plate frame, return them to the plastic pouch with the desiccant pack and seal.
3. Add 100 μ L of **Dilution Buffer** to Blank wells (B2, B3).
4. Add 100 μ L of **Standard** (D2, D3 to G2, G3 and G4, G5 to F4, F5), **sample**, or **positive control** (C2, C3) per well. Cover with plate sealer. Incubate for 2 hours on micro-plate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with **1x Wash Buffer** (300 μ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 μ L of **Detection Antibody working solution** to each well. Cover with plate sealer. Incubate for 2 hours on micro-plate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.

8. Add 100 μ L of **Streptavidin-HRP Conjugate working solution** to each well. Incubate for 60 minutes on micro-plate shaker at room temperature. **Protect from light.**
9. Repeat the aspiration/wash as in step 5.
10. Add 100 μ L of **Substrate Solution** to each well. Incubate for 8-12 minutes on micro-plate shaker at room temperature. **Protect from light.**
11. Add 100 μ L of **Stop Solution** to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 15 minutes, using a micro-plate reader set to 450 nm.

CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control and sample, and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the CXCL14 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

Calculation of samples with a concentration exceeding that of standard 4000 pg/mL may result in inaccurate, low human CXCL14 levels. Such samples require further external predilution according to expected human CXCL14 values with Dilution Buffer in order to precisely quantify the actual human CXCL14 level.

CALIBRATION

This immunoassay is calibrated against a highly purified recombinant human CXCL14/BRAK.

SPECIFICITY

This assay recognizes both natural and recombinant human CXCL14. The factors listed below were prepared at 50 ng/mL in Dilution Buffer, and assayed for cross reactivity. No significant cross-reactivity or interference was observed.

PROTEIN NAME	CROSS-REACTIVITY
Human CXCL14	100%
Human GRO β	0
Human GRO γ	0

LINEARITY

To assess the linearity of the assay, pooled research Human **EDTA plasma** samples were diluted with Dilution Buffer and assayed.

DILUTION FACTOR	ASSAYED (PG/ML)	FINAL (PG/ML)	RECOVERY (%)
5 X	386.742	1933.71	100
10 X	223.621	2236.21	116
20 X	100.736	2014.72	104

To assess the linearity of the assay, pooled research Human **serum** samples were diluted with Dilution Buffer and assayed.

DILUTION FACTOR	ASSAYED (PG/ML)	FINAL (PG/ML)	RECOVERY (%)
5 X	832.088	4160.44	100
10 X	435.170	4351.70	105
20 X	194.271	3885.42	94

TYPICAL DATA

This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

STANDARD (PG/ML)	AVERAGE O.D. (450NM)
Blank	0 (0.081)
125	0.007
250	0.013
500	0.043
1000	0.212
2000	1.095
4000	3.184

- Lot No.:
- Positive Control:

SUMMARY OF ASSAY PROCEDURE**PREPARE REAGENTS, SAMPLES AND STANDARDS**

Add 100 μ L of **standard, samples, positive control** to each well. Incubate 2 hours on the plate shaker at RT.

Aspirate and wash 4 times.

Add 100 μ L **Detection Antibody working solution** to each well. Incubate 2 hours on the plate shaker at RT.

Aspirate and wash 4 times.

Add 100 μ L **Streptavidin HRP conjugate working solution** to each well. Incubate 60 min on the plate shaker at RT. **Protect from light.**

Aspirate and wash 4 times.

Add 100 μ L **Substrate Solution** to each well. Incubate 8-12 min on the plate shaker. **Protect from light.**

Add 100 μ L **Stop Solution** to each well. Read 450nm within 15 min.